

Review Article

Improving and Standardization in Diagnosis of Pancreatic Cancer

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ABSTRACT

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Most cases of pancreatic cancer is lately diagnosed, the commonest reason behind remains diagnostic configuration and deficient in sign and symptoms. Long term survival of pancreatic cancer leans on a solid procedure for diagnosis which leads to concrete impact regaining a successful treatment. As known the current mortality of pancreatic cancer is very close to the incidence. In recent years, progression have been made in diagnosis along with advancement of investigating modalities monitoring and screening of high risk group conversant with exploring wide range of non-invasive tumor markers. Among several strategies, molecular technology has been introduced in diagnosing and prognosis of pancreatic cancer. Lately pancreatic cancer initiation and prognosis including cell cycle, apoptosis, DNA repair, invasivity and metastasis seems to be associated with certain miRNAs. The process may involve positive or negative regulation of expression of proto-oncogenesis and tumor suppressor genes diverse miRNA are expressed abundantly at different level among normal pancreatic tissue, chronic pancreatitis and pancreatic cancer. Hence miRNA serve as tools in differentiating chronic pancreatitis from early stage of cancer. The expectation is high following the early diagnosis of pancreatic cancer. This article is to review the diagnostic framework to build-up the early diagnosis and aimed early treatment of pancreatic cancer.

KEYWORDS: Diagnosis, microRNA, Pancreatic cancer.

INTRODUCTION

Pancreatic cancer (PC) is the 4th most common cause of cancer related death worldwide. 5yrs survival rate is still <5% despite aggressive multidisciplinary treatment and the median survival is 5-8 months.¹ Recent surgical technologies, preoperative management and varieties of investigations have bit improved the conditions of the disease. Development effective of therapies, improvement in survival depends on early diagnosis and treatment modalities based on individual patient characteristics. Therefore early detection of PC is essential to get the satisfied result in curing the disease. However, lack of early diagnosis along with local invasiveness and distant organ metastasis is the prime factor resulting the disease more complicated with poor prognosis.² Risk factors with broad factorial involvement are seen in pancreatic cancer. Familial cancer syndrome and genetic morphological changes have leaded the disease to spread vigorously. Other increased risks of PC are those with breast cancer with fault BRCA2.³ PC also may developed in case of familial adenomatous polyposis (FAP)⁴ and hereditary non polyposis colorectal cancer.5

SCREENING FOR PANCREATIC CANCER: ENDOSCOPY AND ULTRASONOGRAPHY PROCEDURE

a.Endoscopic Retrograde Cholangiopancreatography (ERCP): In Pancreatic malignancies both biliary and pancreatic-duct strictures usually occurs ie "double-duct sign".⁶ ERCP was a gold standard for cystic lesions of the pancreas, and evaluation of biliary strictures or a "double duct sign".7 ERCP biopsy and brush cytology are norms that obtains tissue for diagnosis. However the sensitivity rate for ERCP-directed brush cytology or biopsy is very low.⁸ Techniques to enhance the accuracy of brush cytology, e.g. The technique like digital image analysis, appear to be significantly increased to yield the brush cytology.9 The management of patients with unspecified pancreaticobiliary stricture is a question because of the low sensitivity of current ERCP-guided tissue sampling methods. As finding a proper technique probe-based confocal laser endomicroscopy (pCLE) provides microscopic visualization of strictures along with an ongoing ERCP.¹⁰ The significantly higher accuracy of ERCP and pCLE as compared with ERCP with tissue acquisition, due to its reliable microscopic

examination and its excellent sensitivity and negative predictive value, pCLE is highlighted as modern procedure.¹¹

b. Intraductal Ultrasonography and pancreatoscopy: Pancreatoscopy along with intraductal ultrasonography (IDUS) is used in recent scenario because of its magnificent and reliable diagnostic evaluation instead the single procedure and results shows prominent improvement in differential diagnosis between malignant and benign intraductal papillary mucinous tumor (IPMT). The cumulative survival rate and disease free survival rate are increased by 95% and 93% respectively.¹² Pancreatoscopy provide the direct image structures as a result pancreatic of ductal adenocarcinoma remarkably get differentiated from intraductal papillary mucinous neoplasm and other cystic neoplasm.^{13,14} whereas IDUS exhibit the nature of ductal strictures either benign or malignant.¹⁵

c. Abdominal ultrasound (AUS): AUS represents the minimal invasive and low cost scanning technique easily accessible for monitoring and evaluation of obstructive jaundice and biliary obstruction It may also reveal obvious metastases in liver. Thus, AUS can slightly visualize pancreas in an inadequate fashion, because the deep pancreatic bed and overlapping gas filled stomach or loops of bowel reduced the amplitude of ultrasound. The result differs hand to hand since it has proportionately low sensitivity for spotting small neoplasms within the head of pancreas.¹⁶ So, overcoming the limitations new techniques like color power Doppler ultrasonography, ultrasonographic angiography, contrast harmonic imaging and threedimensional ultrasonography has led to discover the new level of diagnosis improving the sensitivity and specificity of Ultrasonography.^{17,18}

d. Endoscopic ultrasound guided Fine Niddle Aspiration (EUS-FNA): EUS-FNA is widely used in certain circumstances like unresectable pancreatic solid mass. It has a high accurate diagnosis in small mass comparatively with computed tomography (CT) guided FNA.¹⁹ Though it has low predictive value but has relatively high diagnostic accuracy. Because of this drawback it is not recommended in potentially resectable pancreatic tumor however in unresectable pancreatic tumor EUS-FNA is appreciated in need of pathological diagnosis to switch on to neo-adjuvant or palliative radio/chemotherapy. EUS-FNA is a solution to obtain a sample of primary pancreatic lesion or possible metastases which are unsuspected with other imaging modalities.²⁰ Chen et al. demonstrated that EUS was superior to CT and was equivalent to MRI for tumour detection and T and N staging of ampullary tumors.²¹

IMAGING MODALITY FOR PANCREATIC SCREENING

a. Computed tomography (**CT**): Multi-Phase CT scanning has improved the diagnostic capability and has

tremendous efforts in providing high imaging values with more sensitivity and specificity in cancer studies over the past few decades.^{22,23} CT scan is strongly recommended as the primary modality for evaluating patients with suspected malignant biliary obstruction, detection of liver metastases, invasion of vascular structure, potential lymph node involvement including pancreatic neoplasm both for diagnosing and staging.^{24,25} Multidetector computed tomography (MDCT) has improved the accuracy in diagnosis and evaluation of respectability degree of pancreatic neoplasm along with pancreatic parenchymal and peri-pancreatic vascular involvement.²⁶ Based on MDCT, the number of surgery has been doubled with refined techniques combined with new advanced chemo and radiotherapy,²⁷ Neverthless CT and MRI had a low sensitivity (71%) and specificity (58%) in intermediate disease stage ie borderline resectable disease predicting vascular involvement and resectability in the post-chemotherapy.²⁸

b. Magnetic Resonance Imaging (MRI): Multi MRimaging techniques has be evolved a revolution and considered as investigation of choice for diagnosing pancreatic neoplasm.²⁷ Long established MR-imaging yielded high accuracy and less false diagnosis for staging and evaluating pancreatic carcinoma. However, its sensitivity and specificity is still low as compared to CT.8 Gradually to obtain the good result modified technique like Magnetic resonance cholangiopancreatography (MRCP) and Magnetic resonance angiography (MRA) has been discovered. This novel discovery has improved the values of diagnosis and differentiating the degree of staging in pancreatic neoplasm.²⁵ MRCP has elucidate the height and cause of obstruction with strong reliability than CT, recognizing difference between cystic versus solid lesions and also provides excellent ductal imaging, the only drawback is less sensitive in calcified lesions.30 Unenhanced and contrast-enhanced MRI with MRCP and MRA has deliberated considerable remarks in patients suspecting pancreatic tumor.²⁵

c. Positron emission tomography (PET) scan: PETscan has been an investigating modalities of pancreatic cancer in recent days.³² It is proved that F-Fluorodeoxyglucose (FDG) has greater affinity in adenomas which helps to differentiate the nature of the disease.³³ FDG-PET scan is more reliable in pancreatic adenomas rather than pancreatic cancer.³⁴ Similarly in chronic pancreatitis, FDG uptake is lower because of inflammation. However, FDG has excessive avid with salivary gland and suggest that the probability of autoimmune pancreatitis and assumed to recognize pancreatic cancer along with chronic pancreatitis.³⁵ PETscan is considered as better evaluating tools as compared with EUS providing the pooled sensitivity of 90.1% and 81.2% respectively.³⁶ Similarly PET-scan detecting pancreatic cancer has noted the sensitivity of 90% and 95% and specificity of 82% and 100% in similar literature review.^{37,38} For pancreatic cancer staging PETscan has shown similar result as CT and proves that PET-scan has no beneficial effects in recognizing local tumor and regional lymph node spread.³⁹ Similar based imaging 3-deoxy-3 molecular [18F] fluorodeoxyglucose is also in use for differentiating pancreatic cancer. The utility of Flurothymidine-PET (FLT-PET) is narrow in abdominal imaging because of high hepatic uptake⁴⁰ even though some studies demonstrate that FLT-PET is more specific than FDG-PET however the result is vice versa in sensitivity test.⁴¹ The improvement of sensitivity in initial staging of pancreatic cancer has been found by combining FDG-PET along with CT.42

LAPAROSCOPIC STAGING AND LAPAROSCOPIC ULTRASONOGRAPHY

The aim of laproscopic and laproscopic ultrasonography in pancreatic and peri ampullary cancer is sensible to detect the missed occult metastatic lesion in liver and peritoneal cavity where imaging modalities fails to detect the micro lodgement of neoplasms well as neoadjuvant chemo therapy can also be started early because of its reduced invasiveness.⁴³ In case of doubtful MDCT, laproscopic ultrasonography is the investigation of choice and have good approach of detection.⁴⁴ The use of this procedure helps to refine the case of unresectable carcinoma accompanying with an improvement in resection rate.⁴⁵

FECAL DNA AND RNA SCREENING

Blood based markers has been used to approach the noninvasive procedure for distinguishing different cancers, Beside many research had recommended that alteration of genetic and epigenetic changes in RNA and DNA plays a vital role in early detection of cancers and potentially detected in feces in gastrointestinal related cancers.⁴⁶ Secreted pancreatic juice (1.5L/day) flow via bowel and finally excreted in faeces. This fact reflects that molecular changes can also be observed in faecal specimen. As a result faecal biomarkers has been encouraged either for detection of molecular changes in DNA and RNA sequel or simply DNA and RNA Therefore, it has high probability in early detection of Pancreatic cancer.⁴⁷

PANCREATIC JUICE SCREENING

It has been used as alternative biomarkers in early prediction of pancreatic cancer. The several mutation in DNA is observed in the duodenal collection of secretin stimulated pancreatic juice.⁴⁸ This strongly prove that the sample are high quality source that can find a molecular changes in DNA and RNA regarding pancreatic cancer.⁴⁹ Similarly Masao Tanaka et al. reported that twisted expression of RNA in pancreatic juice were hugely degraded into fragments shorter that 200 nucleotides and helps to distinguish pancreatic cancer with non-invasive

neoplasm.⁵⁰ As a result this might improve the early diagnosis and help in upgrading surveillance of patients in pancreatic cancer.

SERUM (BLOOD) TEST FOR THE DETECTION OF PANCREATIC CANCER

Till date there is no commercially recommended food and drug administration (FDA) approved blood test for pancreatic cancer. The majority of blood markers include carcino embryogenic antigen (CEA) and carbohydrate antigens along with majority of protein markers identified by mass spectrometry analysis. New wide range of identification values had raised with detection of molecular changes ie genetic and epigenetic markers (mRNA, DNA, microRNA).^{51,52} The parallel comparison is challenging because of diverse population. As a result wide range of sensitivity and specificity were reported for the various markers. A.K. Siriwardena et al. in a literature review reported, pooled data from 2283 patients evaluated carbohydrate antigen CA19-9, the median sensitivity is 79%(70%-90%) and median specificity 82%(68%-91%) however, specificity of non malignant jaundice in response of CA19-9 is less considerable.53 At present circumstances systematic sample collection, processing and storage should be taken in consideration from large screened population to vield reliable outcomes in early diagnosis of various cancers including pancreatic cancer.

micro RNA

Micro RNAs comprise a novel endogeneous non-coding RNA fragments (22 neuclotide) that plays key role in regulation of gene expression by directing their target mRNA for degradation or translational repression. First microRNA is initially discovered in 1993 by Victor Ambros' in 1993 in Caenorhabditis elegans.⁵⁴ Since then, different mRNA were identified in plants animals and humans. Till the time the updated database listed 2555 humans miRNAs has been explored and most of them are aberrantly expressed in various malignancies.⁵⁵ At present date when we go through human MicroRNAs, disregulation of miRNA are reported in multiple case of cancer and have revealed the clear involvement in disease findings and progression.

miRNA- DYSREGULATION IN TUMORS AND BLOOD SAMPLES

Numerous research groups has compared the status of miRNA in normal pancreatic tissues and pancreatic cancer to analyse the aberrant expression of miRNA similarly large numbers of miRNAs in serum or plasma are reported to be significantly raised and followed by diagnosis of the pancreatic cancer. The several miRNAs are involved and responsible for the transformation of cancers in pancreatic cells. miRNA-21 possess oncogenic effects which are over expressed that increases the proliferation and frequency of cell division in pancreatic cancer.⁵⁶ Similarly, miRNA-221 and

miRNA-192 equally falls on oncogenic genera, over expressed in pancreatic cancer that vulnerably increases the cell cycle progression.⁵⁷ In total evaluation of 29 studies, that reported the status of miRNA in tissue and blood of pancreatic cells are miRNA-21in10 studies, 59,62-67,70,71 miRNA-155 in 7 studies, 63,65,68-70,79,80 miRNA-196a in 12 studies.^{51,58-60,64,67-69,76,79-80}, miRNA-221in 3 studies^{61,68,82} and miRNA-222 in 5 studies,^{60,69,71,80,81} The respective studies showed the aberrant changes in miRNAs. These miRNAs are implicated in development of tumor in pancreatic cells. miRNA-155 has been recently identified as a candidate biomarkers in pancreatic neoplasm. Likewise miRNA-196a has shown the parallel progression of the diseases. The four miRNAs: miRNA-21, miRNA-210, miRNA-155 and miRNA-196a possesses a sensitivity of 64% and specificity of 89%.65 Similarly miRNA-16 and miRNA-196a dominate the independent role in diagnosis of pancreatic cancer, however; these miRNA-16 and miRNA-196a along with combination of CA-19-9 delivered the sensitivity of 92% and specificity of 95.6%.⁵¹ This combination reflects obvious increment in sensitivity and specificity in diagnosis of pancreatic cancer. The evidence strongly supports the facts behind diagnostic characteristics of miRNA in pancreatic cancer.

DISCUSSION

In recent years, large range of evidence has gathered regarding systematisation of clinical investigation related to pancreatic cancer. Some of the investigation has revealed expectations in improving the quality of diagnosis. This review has considered possible available research to generate accuracy in diagnosis of pancreatic cancer. Regardless ERCP - guided brush cytology is standard investigation in pancreatic cancer however, the significance is limited so pCLE (probe-based confocal laser endomicroscopy) is more reliable as modern procedure. Similarly, pancreatoscopy along with intraductal US (IDUS) carries an important role to manifest the nature of ductal strictures to rule out benign or malignant. Plain abdominal ultrasound is not reliable nevertheless Doppler ultrasonography or ultrasonographic angiogram contrast, harmonic imaging and three dimensional ultrasonography has confounding expectation in diagnosis of pancreatic cancer. In case of unresectable pancreatic solid mass FUS-FNA has high accuracy compared to CT. Imaging and occupies broad circumference in diagnosing various solid masses. CT, MDCT, MRI and PET-Scan has improved the accuracy in the field of diagnosis in pancreatic cancer Laparoscopic ultrasonagraphy boost to detect the microlodgement of neoplasm and clears the criteria in resection of carcinoma. Since the recent discovery the role of DNA and microRNA in cellular activity is being observed very closely. The ideas and knowledge on their activities has endeavoured to improvise the technique regarding various carcinomas in the last decade. The divergent physiological process linked with initiation and development of various solid and nonsolid cancers, we have been able to establish the role of miRNA in diagnosing and progression of the disease. Several miRNA like miRNA-21, miRNA155, miRNA-196a, miRNA-221, miRNA-222 are either tumor suppressor or oncogenic molecules respectively, have been illustrated and being identified their role in early detection, prognosis and suitable therapy of the patients. Likewise combination of miRNA-16 and miRNA196a along with CA-19-9 has shown the tremendous result in early diagnosis of pancreatic cancer. We are more optimistic in the indication of miRNA research promising for the use of diagnosing and delivering the personalized medicine to introspect the overall treatment outcome and survival of pancreatic cancer. The success of miRNA seems to have intensed result ever before. The focus should be more determined concerning miRNA in coming future.

ABBREVIATIONS

PC, Pancreatic cancer; FAP, Familial adenomatous polyposis; ERCP, Endoscopic retrograde cholangiopancreatography; pCLE, probe based confocal laser endomicroscopy; IDUS,.Intraductal ultrasonography; IPMT, Intraductal papillary mucinous tumor; EUS-FNA, Endoscopic ultrasound guided fine niddle aspiration; EUS, Endoscopic ultrasonography; CT, Computed tomography; MDCT, Multidetector computed tomography; MRI, Magnetic resonance imaging; MRCP, Magnetic resonance cholangiopancreatography; MRA, Magnetic resonance angiography; PET-Scan, Positron emission tomography FDG, Flurodeoxyglucose; scan; FDG-PET, flurodeocyglucose positron emission tomography; FLT-PET, Flurothymidine positron emission tomography; RNA, Ribonucleic acid; mRNA, messanger RNA; miRNA, microRNA; DNA, Deoxyribonucleic acid; FDA, Food and drug administration; CEA, Carcinoembryonic antigen; CA19-9, Carbohydrate antigen 19-9.

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REFERENCES

1. Siegel RL, Miller KD, Jemal A. Cancer statistics, 2015. CA: a cancer journal for clinicians. 2015;65(1):5-29.

2. Li A, Yu J, Kim H, Wolfgang CL, Canto MI, Hruban RH, et al. MicroRNA array analysis finds elevated serum miR-1290 accurately distinguishes patients with low-stage pancreatic cancer from healthy and disease controls. Clinical Cancer Research. 2013;19(13):3600-10.

3. Luo G, Lu Y, Jin K, Cheng H, Guo M, Liu Z, et al. Pancreatic cancer: BRCA mutation and personalized treatment. Expert Review of Anticancer Therapy. 2015;15(10):1223-31.

4. Mackey R, Walsh RM, Chung R, Brown N, Smith A, Church J, et al. Pancreas-sparing duodenectomy is effective management for familial adenomatous polyposis. Journal of gastrointestinal surgery. 2005;9(8):1088-93.

5. Petersen M, Evert M, Schneider-Stock R, Pross M, Rüschoff J, Roessner A, et al. Serous oligocystic adenoma (SOIA) of the pancreas–first reported case of a genetically fixed association in a patient with hereditary non-polyposis colorectal cancer (HNPCC). Pathology-Research and Practice. 2009;205(11):801-6.

6. Baron TH, Mallery JS, Hirota WK, Goldstein JL, Jacobson BC, Leighton JA, et al. The role of endoscopy in the evaluation and treatment of patients with pancreaticobiliary malignancy. Gastrointestinal endoscopy. 2003;58(5):643-9.

7.Adler D, Schmidt CM, Al-Haddad M, Barthel JS, Ljung B-M, Merchant NB, et al. Clinical evaluation, imaging studies, indications for cytologic study and preprocedural requirements for duct brushing studies and pancreatic fine-needle aspiration: The Papanicolaou Society of Cytopathology Guidelines. CytoJournal. 2014;11(2):1.

8. Mullapudi B, Hawkes PJ, Patel A, Are C, Misra S. Borderline Resectable Pancreatic Cancer. Indian journal of surgical oncology. 2015;6(1):63-8.

9. Kipp BR, Barr Fritcher EG, Pettengill JE, Halling KC, Clayton AC. Improving the accuracy of pancreatobiliary tract cytology with fluorescence in situ hybridization. Cancer cytopathology. 2013;121(11):610-9.

10. Meining A, Chen YK, Pleskow D, Stevens P, Shah RJ, Chuttani R, et al. Direct visualization of indeterminate pancreaticobiliary strictures with probe-based confocal laser endomicroscopy: a multicenter experience. Gastrointestinal endoscopy. 2011;74(5):961-8.

11. Meining A, Shah R, Slivka A, Pleskow D, Chuttani R, Stevens P, et al. Classification of probe-based confocal laser endomicroscopy findings in pancreaticobiliary strictures. Endoscopy. 2012;44(3):251.

12. Hara T, Yamaguchi T, Ishihara T, Tsuyuguchi T, Kondo F, Kato K, et al. Diagnosis and patient management of intraductal papillary-mucinous tumor of the pancreas by using peroral pancreatoscopy and intraductal ultrasonography. Gastroenterology. 2002;122(1):34-43.

13. Itoi T, Sofuni A, Itokawa F, Kurihara T, Tsuchiya T, Ishii K, et al. Initial experience of peroral pancreatoscopy combined with narrow-band imaging in the diagnosis of intraductal papillary mucinous neoplasms of the pancreas (with videos). Gastrointestinal endoscopy. 2007;66(4):793-7.

14. Cunningham SC, Hruban RH, Schulick RD. Differentiating intraductal papillary mucinous neoplasms from other pancreatic cystic lesions. World journal of gastrointestinal surgery. 2010;2(10):331.

15. Kundu R, Pleskow D. Clinical application of intraductal ultrasound during endoscopic retrograde cholangiopancreatography. Gastrointestinal endoscopy clinics of North America. 2009;19(4):615-28.

16. Rösch T, Schusdziarra V, Born P, Bautz W, Baumgartner M, Ulm K, et al. Modern imaging methods versus clinical assessment in the evaluation of hospital in-patients with suspected pancreatic disease. The American journal of gastroenterology. 2000;95(9):2261-70.

17. Mateen MA, Muheet KA, Mohan RJ, Rao PN, Majaz HM, Rao GV, et al. Evaluation of ultrasound based acoustic radiation force impulse (ARFI) and eSie touch sonoelastography for diagnosis of inflammatory pancreatic diseases. JOP Journal of the Pancreas. 2012;13(1):36-44.

18. Sharma C, Eltawil KM, Renfrew PD, Walsh MJ, Molinari M. Advances in diagnosis, treatment and palliation of pancreatic carcinoma: 1990-2010. World journal of gastroenterology: WJG. 2011;17(7):867.

19. Howard TJ, Chin AC, Streib EW, Kopecky KK, Wiebke EA. Value of helical computed tomography, angiography, and endoscopic ultrasound in determining resectability of periampullary carcinoma. The American journal of surgery. 1997;174(3):237-41.

20. Dumonceau J, Polkowski M, Larghi A, Vilmann P, Giovannini M, Frossard J, et al. Indications, results, and clinical impact of endoscopic ultrasound (EUS)-guided sampling in gastroenterology: European Society of Gastrointestinal Endoscopy (ESGE) Clinical Guideline. Endoscopy. 2011;43(10):897.

21. Chen CH, Yang CC, Yeh YH, Chou DA, Nien CK. Reappraisal of endosonography of ampullary tumors: correlation with transabdominal sonography, CT, and MRI. Journal of Clinical Ultrasound. 2009;37(1):18-25.

22. Bronstein YL, Loyer EM, Kaur H, Choi H, David C, DuBrow RA, et al. Detection of small pancreatic tumors with multiphasic helical CT. American Journal of Roentgenology. 2004;182(3):619-23.

23. Fusari M, Maurea S, Imbriaco M, Mollica C, Avitabile G, Soscia F, et al. Comparison between multislice CT and MR imaging in the diagnostic evaluation of patients with pancreatic masses. La radiologia medica. 2010;115(3):453-66.

24. Graf O, Boland G, Warshaw A, Fernandez-del-Castillo C, Hahn P, Mueller P. Arterial versus portal venous helical CT for revealing pancreatic adenocarcinoma: conspicuity of tumor and critical vascular anatomy. AJR American journal of roentgenology. 1997;169(1):119-23.

25. Zhao W-Y, Luo M, Sun Y-W, Xu Q, Chen W, Zhao G, et al. Computed tomography in diagnosing vascular invasion in pancreatic and periampullary cancers: a systematic review and meta-analysis. Hepatobiliary Pancreat Dis Int. 2009;8(5):457-64.

26. Imbriaco M, Megibow AJ, Ragozzino A, Liuzzi R, Mainenti P, Bortone S, et al. Value of the single-phase technique in MDCT assessment of pancreatic tumors. American Journal of Roentgenology. 2005;184(4):1111-7.

27. Shrikhande SV, Arya S, Barreto SG, Ingle S, D'Souza MA, Hawaldar R, et al. Borderline resectable pancreatic tumors: Is there a need for further refinement of this stage? Hepatobiliary & Pancreatic Diseases International. 2011;10(3):319-24.

28. Donahue TR, Isacoff WH, Hines OJ, Tomlinson JS, Farrell JJ, Bhat YM, et al. Downstaging chemotherapy and alteration in the classic computed tomography/magnetic resonance imaging signs of vascular involvement in patients with pancreaticobiliary malignant tumors: influence on patient selection for surgery. Archives of Surgery. 2011;146(7):836-43.

29. Varadhachary GR, Tamm EP, Abbruzzese JL, Xiong HQ, Crane CH, Wang H, et al. Borderline resectable pancreatic cancer: definitions, management, and role of preoperative therapy. Annals of surgical oncology. 2006;13(8):1035-46. 30. Zhang Y, Huang J, Chen M, Jiao LR. Preoperative vascular

evaluation with computed tomography and magnetic resonance imaging for pancreatic cancer: A meta-analysis. Pancreatology. 2012;12(3):227-33.

31. Kim SH, Lee JM, Lee ES, Baek JH, Kim JH, Han JK, Choi BI. Intraductal Papillary Mucinous Neoplasms of the Pancreas: Evaluation of Malignant Potential and Surgical Resectability by Using MR Imaging with MR Cholangiography. Radiology. 2015 Mar;274(3):723-33.

32. Lan BY, Kwee SA, Wong LL. Positron emission tomography in hepatobiliary and pancreatic malignancies: a review. The American Journal of Surgery. 2012;204(2):232-41. 33. Khawandanah MO, Kurkjian C, Penaroza S, Arnold C, Herman TS, Talbert M, Postier R, Pant S. Utility of [18F] flourodeoxyglucose positron emission tomography (FDG-PET) to predict resectability after neoadjuvant therapy in patients with unresectable pancreatic cancer on CT scans. In Journal Of Clinical Oncology 2014 Jan 20 (Vol. 32, No. 3). 2318 Mill Road, Ste 800, Alexandria, Va 22314 Usa: Amer Soc Clinical Oncology.

34. Otomi Y, Otsuka H, Terazawa K, Nose H, Kubo M, Matsuzaki K, et al. Comparing the performance of visual estimation and standard uptake value of F-18 fluorodeoxyglucose positron emission tomography/computed tomography for detecting malignancy in pancreatic tumors other than invasive ductal carcinoma. The Journal of Medical Investigation. 2014;61(1.2):171-9.

35. Deng S-m, Zhang W, Zhang B, Chen Y-y, Li J-h, Wu Y-w. Correlation between the Uptake of 18 F-Fluorodeoxyglucose (18 F-FDG) and the Expression of Proliferation-Associated Antigen Ki-67 in Cancer Patients: A Meta-Analysis. PloS one. 2015;10(6):e0129028.

36. Tang S, Huang G, Liu J, Liu T, Treven L, Song S, Zhang C, Pan L, Zhang T. Usefulness of 18 F-FDG PET, combined FDG-PET/CT and EUS in diagnosing primary pancreatic carcinoma: A meta-analysis. European journal of radiology. 2011 Apr 30;78(1):142-50.

37. Ell PJ. The contribution of PET/CT to improved patient management. The British journal of radiology. 2006 Jan;79(937):32.

38. Gambhir SS, Czernin J, Schwimmer J, Silverman DH, Coleman RE, Phelps ME. A tabulated summary of the FDG PET literature. Journal of nuclear medicine. 2001;42(5 suppl):1S-93S.

39. Kauhanen SP, Komar G, Seppänen MP, Dean KI, Minn HR, Kajander SA, et al. A prospective diagnostic accuracy study of 18F-fluorodeoxyglucose positron emission tomography/computed tomography, multidetector row computed tomography, and magnetic resonance imaging in primary diagnosis and staging of pancreatic cancer. Annals of surgery. 2009;250(6):957-63.

40. Challapalli A, Barwick T, Pearson RA, Merchant S, Mauri F, Howell EC, et al. 3'-Deoxy-3'-18F-fluorothymidine positron emission tomography as an early predictor of disease progression in patients with advanced and metastatic pancreatic cancer. European journal of nuclear medicine and molecular imaging. 2015;42(6):831-40.

41. Herrmann K, Erkan M, Dobritz M, Schuster T, Siveke J, Beer A, et al. Comparison of 3'-deoxy-3'-[18F] fluorothymidine positron emission tomography (FLT PET) and FDG PET/CT for the detection and characterization of pancreatic tumours. European journal of nuclear medicine and molecular imaging. 2012;39(5):846-51. 42. Farma JM, Santillan AA, Melis M, Walters J, Belinc D, Chen D-T, et al. PET/CT fusion scan enhances CT staging in patients with pancreatic neoplasms. Annals of surgical oncology. 2008;15(9):2465-71.

43. Giovanni D. Single-Incision Laparoscopic Gastrojejunal Bypass with Intraoperative Ultrasonography for Obstructive Pancreatic Head Tumor. Journal of Laparoendoscopic & Advanced Surgical Techniques. 2014;24(4).

44. Barabino M, Santambrogio R, Ceretti AP, Scalzone R, Montorsi M, Opocher E. Is there still a role for laparoscopy combined with laparoscopic ultrasonography in the staging of pancreatic cancer? Surgical endoscopy. 2011;25(1):160-5.

45. Hariharan D, Constantinides V, Froeling F, Tekkis P, Kocher H. The role of laparoscopy and laparoscopic ultrasound in the preoperative staging of pancreatico-biliary cancers–a meta-analysis. European Journal of Surgical Oncology (EJSO). 2010;36(10):941-8.

46. Wu CW, Ng SS, Dong YJ, Ng SC, Leung WW, Lee CW, et al. Detection of miR-92a and miR-21 in stool samples as potential screening biomarkers for colorectal cancer and polyps. Gut. 2011:gut. 2011.239236.

47. Bhat K, Wang F, Ma Q, Li Q, Mallik S, Hsieh T-c, et al. Advances in biomarker research for pancreatic cancer. Current pharmaceutical design. 2012;18(17):2439.

48. Dal Molin M, Matthaei H, Wu J, Blackford A, Debeljak M, Rezaee N, et al. Clinicopathological correlates of activating GNAS mutations in intraductal papillary mucinous neoplasm (IPMN) of the pancreas. Annals of surgical oncology. 2013;20(12):3802-8.

49. Kanda M, Sadakari Y, Borges M, Topazian M, Farrell J, Syngal S, et al. Mutant TP53 in duodenal samples of pancreatic juice from patients with pancreatic cancer or high-grade dysplasia. Clinical Gastroenterology and Hepatology. 2013;11(6):719-30. e5.

50. Ohuchida K, Mizumoto K, Ohhashi S, Yamaguchi H, Konomi H, Nagai E, et al. Twist, a novel oncogene, is upregulated in pancreatic cancer: clinical implication of Twist expression in pancreatic juice. International journal of cancer. 2007;120(8):1634-40.

51. Liu J, Gao J, Du Y, Li Z, Ren Y, Gu J, et al. Combination of plasma microRNAs with serum CA19-9 for early detection of pancreatic cancer. International Journal of Cancer. 2012;131(3):683-91.

52. Richards EJ, Kong W, Malafa M, Cheng JQ, Coppola D. Molecular Diagnostics of Pancreatic Cancer. Molecular Pathology and Diagnostics of Cancer: Springer; 2014. p. 259-82.

53. Goonetilleke K, Siriwardena A. Systematic review of carbohydrate antigen (CA 19-9) as a biochemical marker in the diagnosis of pancreatic cancer. European Journal of Surgical Oncology (EJSO). 2007;33(3):266-70.

54. Ambros V, Horvitz H. Heterochronic mutants of the nematode. Science. 1984;226:409-16.

55. Fazlul, HS. Evolving concept of cancer stem cells: Role of micro-RNAs and their implications in tumor aggressiveness. Journal of Carcinogenesis & Mutagenesis. 2011 Jun 17.

56. Park J-K, Lee EJ, Esau C, Schmittgen TD. Antisense inhibition of microRNA-21 or-221 arrests cell cycle, induces apoptosis, and sensitizes the effects of gemcitabine in pancreatic adenocarcinoma. Pancreas. 2009;38(7):e190-e9.

57. Zhao C, Zhang J, Zhang S, Yu D, Chen Y, Liu Q, et al. Diagnostic and biological significance of microRNA-192 in

pancreatic ductal adenocarcinoma. Oncology reports. 2013;30(1):276-84.

58. Slater EP, Strauch K, Rospleszcz S, Ramaswamy A, Esposito I, Klöppel G, et al. MicroRNA-196a and-196b as potential biomarkers for the early detection of familial pancreatic cancer. Translational oncology. 2014;7(4):464-71.

59. Wang J, Chen J, Chang P, LeBlanc A, Li D, Abbruzzesse JL, et al. MicroRNAs in plasma of pancreatic ductal adenocarcinoma patients as novel blood-based biomarkers of disease. Cancer prevention research. 2009;2(9):807-13.

60. Zhang Y, Li M, Wang H, Fisher WE, Lin PH, Yao Q, et al. Profiling of 95 microRNAs in pancreatic cancer cell lines and surgical specimens by real-time PCR analysis. World journal of surgery. 2009;33(4):698-709.

61. Lee EJ, Gusev Y, Jiang J, Nuovo GJ, Lerner MR, Frankel WL, et al. Expression profiling identifies microRNA signature in pancreatic cancer. International journal of cancer. 2007;120(5):1046-54.

62. Dillhoff M, Liu J, Frankel W, Croce C, Bloomston M. MicroRNA-21 is overexpressed in pancreatic cancer and a potential predictor of survival. Journal of Gastrointestinal Surgery. 2008;12(12):2171-6.

63. Bhatti I, Lee A, James V, Hall RI, Lund JN, Tufarelli C, et al. Knockdown of microRNA-21 inhibits proliferation and increases cell death by targeting programmed cell death 4 (PDCD4) in pancreatic ductal adenocarcinoma. Journal of Gastrointestinal Surgery. 2011;15(1):199-208.

64. Du Rieu MC, Torrisani J, Selves J, Al Saati T, Souque A, Dufresne M, et al. MicroRNA-21 is induced early in pancreatic ductal adenocarcinoma precursor lesions. Clinical chemistry. 2010;56(4):603-12.

65. Bauer AS, Keller A, Costello E, Greenhalf W, Bier M, Borries A, et al. Diagnosis of pancreatic ductal adenocarcinoma and chronic pancreatitis by measurement of microRNA abundance in blood and tissue. PLoS One. 2012;7(4):e34151.

66. Liu R, Chen X, Du Y, Yao W, Shen L, Wang C, et al. Serum microRNA expression profile as a biomarker in the diagnosis and prognosis of pancreatic cancer. Clinical chemistry. 2012;58(3):610-8.

67. Kong X, Du Y, Wang G, Gao J, Gong Y, Li L, et al. Detection of differentially expressed microRNAs in serum of pancreatic ductal adenocarcinoma patients: miR-196a could be a potential marker for poor prognosis. Digestive diseases and sciences. 2011;56(2):602-9.

68. Bloomston M, Frankel WL, Petrocca F, Volinia S, Alder H, Hagan JP, et al. MicroRNA expression patterns to differentiate pancreatic adenocarcinoma from normal pancreas and chronic pancreatitis. Jama. 2007;297(17):1901-8.

69. Jamieson NB, Morran DC, Morton JP, Ali A, Dickson EJ, Carter CR, et al. MicroRNA molecular profiles associated with diagnosis, clinicopathologic criteria, and overall survival in patients with resectable pancreatic ductal adenocarcinoma. Clinical Cancer Research. 2012;18(2):534-45.

70. Preis M, Gardner TB, Gordon SR, Pipas JM, Mackenzie TA, Klein EE, et al. MicroRNA-10b expression correlates with response to neoadjuvant therapy and survival in pancreatic ductal adenocarcinoma. Clinical Cancer Research. 2011;17(17):5812-21.

71. Park J-K, Henry JC, Jiang J, Esau C, Gusev Y, Lerner MR, et al. miR-132 and miR-212 are increased in pancreatic cancer and target the retinoblastoma tumor suppressor. Biochemical

and biophysical research communications. 2011;406(4):518-23.

72. Zhang S, Hao J, Xie F, Hu X, Liu C, Tong J, Zhou J, Wu J, Shao C. Downregulation of miR-132 by promoter methylation contributes to pancreatic cancer development. Carcinogenesis. 2011 Aug;32(8):1183.

73. Yu S, Lu Z, Liu C, Meng Y, Ma Y, Zhao W, et al. miRNA-96 suppresses KRAS and functions as a tumor suppressor gene in pancreatic cancer. Cancer research. 2010;70(14):6015-25.

74. Vogt M, Munding J, Grüner M, Liffers S-T, Verdoodt B, Hauk J, et al. Frequent concomitant inactivation of miR-34a and miR-34b/c by CpG methylation in colorectal, pancreatic, mammary, ovarian, urothelial, and renal cell carcinomas and soft tissue sarcomas. Virchows Archiv. 2011;458(3):313-22.

75. Torrisani J, Bournet B, Du Rieu MC, Bouisson M, Souque A, Escourrou J, et al. let-7 MicroRNA transfer in pancreatic cancer-derived cells inhibits in vitro cell proliferation but fails to alter tumor progression. Human gene therapy. 2009;20(8):831-44.

76. Ho AS, Huang X, Cao H, Christman-Skieller C, Bennewith K, Le Q-T, et al. Circulating miR-210 as a novel hypoxia marker in pancreatic cancer. Translational oncology. 2010;3(2):109-13.

77. Wang W-S, Liu L-X, Li G-P, Chen Y, Li C-Y, Jin D-Y, et al. Combined serum CA19-9 and miR-27a-3p in peripheral blood mononuclear cells to diagnose pancreatic cancer. Cancer Prevention Research. 2013;6(4):331-8.

78. Greither T, Grochola LF, Udelnow A, Lautenschläger C, Würl P, Taubert H. Elevated expression of microRNAs 155, 203, 210 and 222 in pancreatic tumors is associated with poorer survival. International Journal of Cancer. 2010;126(1):73-80.

79. Szafranska A, Davison T, John J, Cannon T, Sipos B, Maghnouj A, et al. MicroRNA expression alterations are linked to tumorigenesis and non-neoplastic processes in pancreatic ductal adenocarcinoma. Oncogene. 2007;26(30):4442-52.

80. Schultz NA, Werner J, Willenbrock H, Roslind A, Giese N, Horn T, et al. MicroRNA expression profiles associated with pancreatic adenocarcinoma and ampullary adenocarcinoma. Modern Pathology. 2012;25(12):1609-22.

81. Szafranska AE, Doleshal M, Edmunds HS, Gordon S, Luttges J, Munding JB, et al. Analysis of microRNAs in pancreatic fine-needle aspirates can classify benign and malignant tissues. Clinical chemistry. 2008;54(10):1716-24.

82. Kawaguchi T, Komatsu S, Ichikawa D, Morimura R, Tsujiura M, Konishi H, et al. Clinical impact of circulating miR-221 in plasma of patients with pancreatic cancer. British journal of cancer. 2013;108(2):361-9.

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